cDNA Cloning and Gene Expression of Anthocyanidin Synthase from Torenia fournieri

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Abstract

The cDNA clone, TAN1, encoding anthocyanidin synthase (ANS) was isolated by screening of a cDNA library constructed from flower petals of *Torenia fournieri* cv. Summer Wave Blue using a heterologous probe. Nucleotide sequence analysis revealed that it contains an open reading frame encoding a polypeptide of 376 amino acids. The postulated amino acid sequence shows 48 % to 72 % identities with those of previously reported ANSs. Southern blot analysis suggested that there is a single copy of *ANS* gene in the genome of *T. fournieri*. Northern blot analysis indicated that *ANS* gene is expressed in flower bud but not in leaf or mature flower.

The nucleotide sequence reported in this paper has been deposited in GenBank/EMBL/DDBJ databases with accession number AB044091.

Anthocyanins, representative secondary metabolite found in most plant species, are formed through flavonoid biosynthetic pathway (Heller and Forkmann, 1993). Enzymes responsible for most steps of the pathway have been described, and their cDNAs have been cloned from various plant species (Dooner and Robbins, 1991; Forkmann, 1993; Holton and Cornish, 1995; Martin and Gerats, 1992). Anthocyanidin synthase (ANS) catalyzes the reaction leading to anthocyanidin from leucoanthocyanidin, the first step for the formation of colored metabolites, and thus an important step in anthocyanin formation. In our previous paper (Saito et al., 1999), we studied the mechanism of ANS reaction catalyzed by the recombinant enzyme from Perilla frutescens. In the present study, we have isolated a cDNA encoding ANS from the flowers of Torenia fournieri, and compared with ANS proteins from various plant species. T. fournieri is a commercial ornamental plant and also a suitable experimental model for genetic engineering of flower color (Suzuki et al., in press).

The ANS cDNA, *TAN1*, was isolated from *T. fournieri* cv. Summer Wave Blue (Suntory Ltd.) by screening of λ ZAP II cDNA library (Stratagene) constructed from flower petals using the heterologous probe isolated from *Perilla frutescens* (Saito

et al., 1999). Nucleotide sequence analysis was carried out by dideoxy chain termination method using automatic sequencer (model DSQ-2000L, Shimadzu, Japan). The nucleotide sequence of TAN1 was 1348 bp long, and contained open reading frame of 1128 bp encoding 376 amino acids forming a 42,130-Da polypeptide (Fig. 1). The authenticity of the first ATG codon was confirmed by the presence of an in-frame stop codon (TAA) at 18-bp upstream from the first ATG. Sequence comparison of amino acid sequence of TAN1 with those of reported ANSs indicated 48-72 % identity and 60-80 % similarity in amino acid residues (Fig. 2A). His-240, Asp-242 and His-296 are completely conserved in ANSs from several plant species, which are thought to bind with iron at the catalytic center of the iron-containing soluble oxygenases (Britsch et al., 1993; De Carolis and De Luca, 1994; Matsuda et al., 1997; Prescott and John, 1996; Prescott, 1993). The Arg-Xaa-Ser (306-308) motif, reported as the specific binding site of 2 -oxoglutarate in flavanone 3β -hydroxylase from petunia (Lukacin and Britsch, 1997; Lukacin et al., 2000), is also conserved in ANSs. From phylogenetic tree (Fig. 2B), the Torenia ANS falls into a group of dicot ANSs. The phylogenetic tree also indicated the presence of distinct difference

- 71 CGGCCCGTGTCGAACTACTGTCGAACAGCGGAATCAAAGCGATCCCGAAAGAGTACGTAAGGACCGAAGA A R V E L L S N S G I K A I P K E Y V R T E E
- 141 AGAGCTCCGGAGCATAACCGACATCTTCTCGAAAGAAGATGGCGCGAAAAACATCGACAGCCCAGATTTG E L R S I T D I F S K E D G A K N I D S P D L
- 211 CCCATCATCGACCTATCGAAGATCGACTCGAGCGACGAAGAAGACGCGCAAGAAAGGCCACGAGGAGTTGA P I I D L S K I D S S D E E T R K K G H E E L K
- 281 AAGAAGCAGCGATCGAATGGGGCGTCATGCATCTGATCAACCACGGAATATCGGACGAGCTGATCAATCG E A A I E W G V M H L I N H G I S D E L I N R
- 351 GGTCAAGAAAGCGGGCGGCGAGTTCTTCGATCTCCCCGTCGAGGAGAAGGAGAAGTATGCGAACGACCAG V K K A G G E F F D L P V E E K E K Y A N D Q
- 421 TCGAGCGGGAACGTTCAGGGGTACGGAAGTAAATTGGCGAACAACGCGGGGTGGGATTTTGGAGTGGGAGG S S G N V Q G Y G S K L A N N A G G I L E W E D
- 491 ACTATTTCTTCCATTGCGTGTATCCCCGAGGAGAAGAGGGGATATGGCTATTTGGCCTAAGGATCCACAAGA Y F F H C V Y P E E K R D M A I W P K D P Q D
- 561 TTACATCCCCGCAACGACCGAGTACGCGAAAGAGATACGATCCTCACGACCAAGATCCTCTCGGTCCTC Y I P A T T E Y A K E I R S L T T K I L S V L
- 631 TCGCTCGGCCTCGGGCTCGACCAAGACCGTCTTGAGAAAGAGGGTCGGTGGAAAAGACGACCTCACCCTCC S L G L G L D Q D R L E K E V G G K D D L T L Q
- 701 AAATGAAGATAAACTACTACCCGAAATGCCCTCAACCCGAGCTAGCCCTGGGCGTCGAGGCCCACACGGA M K I N Y Y P K C P Q P E L A L G V E A H T D
- 771 CGTGAGCGCCCTCACGTTCATCCTCCACAACATGGTCCCGGGCCTCCAAGTGCTCTACAAAGGCGAGTGG V S A L T F I L H N M V P G L Q V L Y K G E W
- 841 GTCACCGCTTCATGCGTGCCCGACTCCATAATCTTGCATGTTGGGGACACGATCGAGATCTTGAGCAATG V T A S C V P D S I I L H V G D T I E I L S N G
- 911 GCATGTATAAGAGCGTGCTCCATAGGGGTTTGGTGAATCGGGAGAGGGTTAGGGGTTTCTTGGGCGGTCTT M Y K S V L H R G L V N R E R V R V S W A V F
- 981 TTGCGAGCCGCCAAGGAGAAGATCGTGCTCAAGCCACTGCCCGAGACGGTCGGCGAGGATCGTCCCGCT C E P P K E K I V L K P L P E T V G E D R P A
- 1051 CTGTTTCCGCCTCGGACTTTCGCGCAACACATGAAGCATAAGTTGTTCAAGAAGAGCGATGATGAAGTGC L F P P R T F A Q H M K H K L F K K S D D E V H
- 1191 TIGATGATTGTTTGGATTGGATTGTGAATGTTTGCAATTGTGTTTTTCTGTGATTAATGGATGTATAATGTCCG

1261 GATTATTTTAAGAATATATAACCGTATGTTTTATGTACCATGAAAAATAAAGTAATGCTTTTAATTGGCT

- 1331 AAAAAAAAAAAAAAAAAAA 1348
- Fig. 1. Nucleotide sequence and deduced amino acid sequence of Torenia ANS. Asterisk indicates in-flame stop codon.

between the sequences of monocotyledoneae and dicotyledoneae.

For Southern blot analysis, 2.5 μ g of genomic DNA from leaves of *T. fournieri* isolated by the described method (Murray *et al.*, 1980) was digested with several different restriction enzymes, which do not cut within the *ANS* cDNAs. Then the digested DNAs were separated by electrophoresis on a 0.8 % agarose gel, transferred onto HybondN+ membrane (Amersham) and hybridized with a ${}^{32}P$ -labeled cDNA probe at 60 °C. As a probe, coding region of *ANS* cDNA was labeled by ${}^{32}P$ -dCTP using Random Primer DNA Labeling Kit (Takara Shuzo, Japan) and purified by NAP5 Column (Pharmacia Biotech). The membrane was finally washed in 0.1 × SSPE, 0.1 % SDS at 60 °C for 10 min. Hybridization signals were detected with a BAS-2000 Image Analyzer (Fuji Film,



torenia snapdragon perilla petunia grape forsythia	11MVSPASPSPARVELLINGGINATPKEY REEELRSITO FSKEDGAKNI SED PIIDISKIDSSEIETEKEGHEELKEAA WGVMHL NH; 11MVTSAMGPSPRVELL KSGIGSIPKEY REEELRSIGD FAEERNN GPC PVIDLAAINS DEEVRKCH ELKKAA WGVMHL NH; 11MVTSAMGPSPRVELARSG DIFFK Y REEELRSIG FAEERSG GPC PVIDLAAINS DEEVRKCHEELKKAAI WGVMHL NH; 11MVTSAMGPSPRVELARSG DIFFK Y REBELRSIG FIEERSG GPC PVIDLAAINS DEEVRKCHEELKKAAI WGVMHL NH; 11MVTSVAPRVESLARSG DIFFK Y REBELRSIG FIEERKD GPC PVIDLKEDS DKEIRKCHEELKKAAI WGVMHL NH; 11MVTSVAPRVESLARSG DIFFK Y RECENTSIG FIEEKD GPC PVIDLKEDS DKEIRKCHEELKKAA WGVMHL NH; 11MVTSVAPRVESLASGIGJFFKFY RFCENTSIG FIEEKD GPC PVIDLKEDS DEVWR.EIRE GREELKKAA WGVMHL NH; 11NVTSVAPRVESLASGIGSIFKFY RFCENTSIG FIEEMSD GPC PVIDLKEDS DEVWR.EIRE GREELKKAA WGVMHL NH;	93 89 91 89 91 87
apple carrot chinaaster arabidopsis commonstock sweetpotato maize rice	1WYSYDSYNSRYETLAGSGIGITPREY REX EUWHGD FE EKING GFO PTIDIKETS EKVRAKGREGKAA WGYMHL NH 1WYTTLESFNETLAGSGIGLTPREY REX ELESTT PEEL.KED GFO PTIDIKETS EKVRAKGREGKAA WGYMHL NH 1WYTTLESFNETRYESIAN SGIDIFREY REX ELESTT PDEL.KED GFO PTIDIKETS DEAVREKYEGL KAAM WGYMHL NH 1MYAVENVESIAN SGIDIFREY REX ELESTT PDEL.KED GFO PTIDIKETS DEAVREKYEGL KAAM WGYMHL NH 1MYAVENVESIAN SGIDIFREY REX ELESTT PDEL.KED GFO PTIDIKETS DEATREKYEGL KAAM WGYMHL NH 1MYAVENVESIAN SGIDIFREY REX ELESTT PDEL.KET GFO PTIDIKETS DEATREKYEGL KAAM WGYMHL NH 1MYAVENVESIAN SGIDIFREY REX ELESTT PDEL KAEK GFO PTIDIKETS DEATREKYEGL KAAM WGYMHL NH 1MYAVENVESIAN SGIDIFREY REX ELESTT PDEL KAEK GFO PTIDIKETS DEATREKYEGL KAAM WGYMHL NH 1MYAVENVESIAN SGISTPKEY REX ELESTT PDELSIND FOEL.KEEK GFO PTIDIKETS DEATREKYEGL KAAM WGYMHL NH 1MYAVENVESIAN SGISTPKEY REX ELESTT PALEN KEK GFO PTIDIKETS DEATREKYEGL KAAM WGYMHL NH 1	89 89 87 85 85 91 98 89
torenia snapdragom perilla petunia grape forsythia apple carrot chinaster arabidopsis commonstock sweetpotato maize rice	94 GISDEL NAVKAAGGEFFD LEVEEKEKYANDO GN GGYGSKLANNAGGI LEWEDYFFEC V FE KRO IVEEKO DY FAT TYAK E RSUTTK 19: 90 G PDELTANYKAGOEFFD LEVEKEKIANDOA GN GGYGSKLANNASGLEWEDYFFEC V FE KRO IVEEKEFD VI PAT BYAK E RSUTTK 19: 92 GIPDELTANYKAGOEFFD LEVEKEKIANDOA GN GGYGSKLANNASGLEWEDYFFEC VE KRO IVER PDVI PAT BYAK E RSUTTK 19: 90 GISDEL NAVKAGIFFTD OP VEEKEKVANDOA GN GGYGSKLANNASGLEWEDYFFEC VE KRO IVER PDVI PAT BYAK E RSUTK 19: 90 GISDEL NAVKAGIFFTD OP VEEKEKVANDOA GN GGYGSKLANNASGLEWEDYFFEC VE KROISIWER PDVI PAT BYAK GRADATK LT 88: GISDEL DAVKAGIFTSD DE EKEKVANDOA GN GGYGSKLANNASGLEWEDYFFEC VE KROISIWER PTDVI PAT BYAK GRADATK LT 90 GISDEL DAVKAGIFTSD PEKEKVANDOA GN GGYGSKLANNASGLEWEDYFFEC VE KROISIWER PTDVI PAT BYAK GRADATK LT 88: GISDEL DAVKAGITFTD PEKEKVANDOA GN GGYGSKLANNASGLEWEDYFFEL VE KROISIWER PTDVI PAT BYAK GRADATK LT 90 GISDEL DAVKAGCAFFN GEVEKKVANDOA GN GGYGSKLANNASGLEWEDYFFEL VE KROISIWER PTDVI PAT BYAK GRADATK LT 88: GISDEL DAVKAGCAFFN GEVEKKVANDOA GN GGYGSKLANNASGLEWEDYFFEL VE KROISIWER PTDVI PAT BYAK GRADATK LT 90 GISDEL DAVKAGCAFFN GEVEKKVANDOA GN GGYGSKLANNASGLEWEDYFFEL VE KADISIWER PTDVI PAT BYAK GRADATK LS 86: GISSEL DAVKAGCAFFSD OP VEEKKVANDOA GX GGYGSKLANNASGLEWEDYFFEL PE KADISIWER PTDVI PAT BYAK GRADATK LS 87: GISSEL DAVKAGCAFFSD OP VEEKKVANDOA GX GGYGSKLANNASGLEWEDYFFEL VE FRUISIWER PTDVI PAT BYAK GRADATK LS 86: GIFYL LERKKKAGEFFSLEVYANDOA GX GGYGSKLANNASGLEWEDYFFEL VE FRUISIWER PTDVI PAT BYAK GRADATK LS 86: GIFYL LERKKKAGEFFSLEVYANDOA GX GGYGSKLANNASGLEWEDYFFEL PFEKTLIS WER PTDVI FAT BYAK GRADATK LS 86: GIFYL LERKKKAGEFFSLEVYANDOA GX GGYGSKLANNASGLEWEDYFFEL VE FRUISIWER PTDVI FAT BYAK GRADATK CR 86: GIFYL LERKKKAGEFFSLEVYANDOA GX GGYGSKLANNASGLEWEDYFFEL VE PE KROLSIWER PTDVI FAT BYAK GRADATK CR 86: GIFYL LERKKKAGEFFFELVYANDOA GX GGYGSKLANNASGLEWEDYFFEL VE PE KROLSIWER PTDVI FAT BYAK GRADATK CR 86: GIFYL LERKKKAGEFFFELVYANDOA GX GYGSKLANNASGLEWEDYFFEL VE PE KROLSIWER PTDVI FAT BYAK GRADATK CR 86: GIFYL LERKKKAGEFFFELVYANDOA GX GGYGSKLANNASGLEWEDYFFEL VE PE KROLSIWER PTDVI FAT BYAK GRADATK CR 87 0 GR AA	193 189 191 189 191 186 189 185 185 185 198 198
torenia snapdragon perilla petunia grape forsythia apple carrot chinasater arabidopsis commonatock sweetpotato maize rice	1941 LS GLG. LS OL RLEKEVGGK L LLONKINY YPKCPOPELALGVEAHTDVSALTFILHNNVFGLONDVKGVPDGII H.GDT G 1900 LS GLG. LSPERLEKEVGG E L LONKINY YPKCPOPELALGVEAHTDVSALTFILHNNVFGLOF YRGKWVTAKCVPNGIIMH GDT G 1901 LS GLG. LSFOR LSKEVGG E LLONKINY YPKCPOPELALGVEAHTDVSALTFILHNNVFGLOF YRGKWVTAKCVPNGIIMH GDT G 1902 LS GLG. LSFOR LSKEVGG E LLONKINY YPKCPOPELALGVEAHTDVSALTFILHNNVFGLOF YRGKWVTAKCVPNGIIMH GDT G 1903 LS GLG. LSFOR LSKEVGG E LLONKINY YPKCPOPELALGVEAHTDVSALTFILHNNVFGLOF YRGKWVTAKCVPNGIIMH GDT G 1904 LSGLG. LSFOR LSKEVGG E LLONKINY YPKCPOPELALGVEAHTDVSALTFILHNNVFGLOF YRGKWVTAKCVPNGIIMH GDT G 1905 LSGLG. LSFOR LSKEVGG E LLONKINY YPKCPOPELALGVEAHTDVSALTFILHNNVFGLOF YRGKWVTAKCVPNGIIMH GDT G 1904 LSGLG. LSFOR LSKEVGG E LLONKINY YPKCPOPELALGVEAHTDVSALTFILHNNVFGLOF YRGKWVTAKCVPNGIIMH GDT G 1904 LSGLG. LSFOR LSKEVGG E LLONKINY YPKCPOPELALGVEAHTDVSALTFILHNNVFGLOF YGKWVTAKCVPNGIIMH GDT G 1906 LSGLG. LSFOR LSKEVGG E LLONKINY YPKCPOPELALGVEAHTDVSALTFILHNNVFGLOF YGKWVTAKCVPNGIIMH GDT G 1906 LSGLG. LSFOR LSKEVGG E LLONKINY YPKCPOPELALGVEAHTDVSALTFILHNNVFGLOF YGKWVTAKCVPDSII MH GDT G 1906 LSGLG. LSFOR LSKEVGG E LLONKINY YPKCPOPELALGVEAHTDVSALTFILHNNVFGLOF YGKWVTAKCVPDSII MH GDT G 1907 LSGLG. LSFOR LSKEVGG E LLONKINY YPKCPOPELALGVEAHTDVSALTFILHNNVFGLOF YGKWVTAKCVPDSI MH GDT G 1909 LSGLG. LSFOR LSKEVGG B LLONKINY YPKCPOPELALGVEAHTDVSALTFILHNNVFGLOF YGKWVTAKCVPDSI MH GDT G 1909 LSGLG. LSFOR LSKEVGG B LLONKINY YPKCPOPELALGVEAHTDVSALTFILHNNVFGLOF YGKWVTAKCVPDSI MH GDT G 1909 LSGLG. LSFOR LSKEVGG B LLLONKINY YPKCPOPELALGVEAHTDVSALTFILHNNVFGLOF YGKWVTAKCVPDSI MH GDT G 1909 LSGLG. LSFOR LSKEVGG B LLLONKINY YPKCPOPELALGVEAHTDVSALTFILHNNVFGLOF YGKWVTAKCVPDSI MH GDT G 1909 LSGLG. LSFOR LSKEVGG B LLLONKINY YPKCPOPELALGVEAHTDVSALTFILHNNVFGLOF YGKWVTAKCVPDSI MH GDT G 1909 LSGLG. LSFOR LSKEVGG B LLLONKINY YPKCPOPELALGVEAHTDVSALTFILHNNVFGLOF YGKWVTAKCVPDSI MH GDT G 1909 LSGLG. LSFOR LSKEVGG B LLLONKINY YPKCPOPELALGVEAHTDVSALTFILHNNVFGLOF YGKWVTAKCVPDSI MH GDT G 1909 LSGLGLG. LSFOR LSKEVGG B LLLONKINY YPCCPOPELALGVEAHTDVSALTFILHNNVF	284 280 282 280 282 277 280 278 276 276 276 298 2986
torenia snapdragon perilla petunia grape forsythia apple carrot chinaaster arabidopsis commonatock sweetpotato mize rice	185: ILSNGMYKS LHRGLVN E VR SWAVFCEPPKEKI LKFLPETVGEDR. FALFPERTFAGH KHLFR SODEVHDEGUVVEHDESVE. 181: LSNGKYKSILHRGLVNKEKVRISWAVFGEPKEKI LKFLPETV EV. FAFAGH KHLFR SODEVHDEGUVVEHDESVE. 181: LSNGKYKSILHRGLVNKEKVRISWAVFGEPKEKI LØFLETV EV. FFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFF	375 373 362 379 362 357 355 3556 3556 3955 375
torenia snapdragon perilla petunia grape forsythia apple carrot chinaaster arabidopsis commonstock sweetpotato maize rice	380: QDAVAENKDIKEDEQCGPAEHKDIKEDGQGAAAENKVFKENNQDVAAEESK 430 380: KTEESNPQKIEILKPGEEASSSP	
(B)	maize rice	
	torenia	
	snapdragon	
c	rabidopsis	
	carrot forsythia	
	appie / petunia chinaaster	
	grape	

Fig. 2. Multiple alignment (A) and phylogenetic tree (B) of deduced amino acid sequences of ANSs from several plant species. (Continued on back page)

Multiple alignment of amino acid sequences was carried out by clustal W program (Thomopson et al., 1994). Black shading indicates identical amino acids and gray shading shows similar amino acids. Dashes indicate gaps in the sequences for best alignment. Arrowheads show conserved His and Asp residues postulated as required for ferrous-ion coordination. Asterisks indicate the Arg-Xaa-Ser motif conserved in flavanone 3β -hydroxylase from petunia, assumed to be a binding site of 2-oxoglutarate (Lukacin and Britsch, 1997; Lukacin et al., 2000). Phylogenetic tree was generated by neighbor - joining method (Saitou and Nei, 1987). torenia, Torenia fournieri (this study); snapdragon, Antirrhinum majus (Martin et al., 1991); perilla, Perilla frutescens (Saito et al., 1999); petunia, Petunia hybrida (Weiss et al., 1993); grape, Vitis vinifera L. (Sparvoli et al., 1994); forsythia, Forsythia x intermedia (Rosati et al., 1999); apple, Malus sp. (Davies, 1993); carrot, Daucus carota (Hirner and Seitz, accession number AF184273); chinaaster, Callistephus chinensis (Henkel and Forkmann, accession number AF015885); arabidopsis, Arabidopsis thaliana (Pelletier et al., 1997); commonstock, Matthiola incana (Min and Forkmann, accession number AF026058); sweetpotato, Ipomoea batatas (Shiokawa et al., accession number AB023786); maize, Zea mays (Messen et al., 1990); rice, Oryza sativa (Reddy, accession number Y07955).



Fig. 3. Southern blot (A) and northern blot (B) analyses of ANS gene from Torenia fournieri. (A) Genomic DNA ($2.5 \ \mu g$) isolated from leaves were digested with EcoRI, EcoRV, HindIII and PstI, separated on an agarose gel ($0.8 \ \%$), transferred onto a membrane, and then hybridized with ³²P-labeled TAN1 coding region as the probe. (B) Total RNA ($10 \ \mu g$) isolated from petals of flower buds, mature flower and leaves were electrophoresed on an agarose gel ($1.2 \ \%$), transferred to a membrane, and then hybridized with the probe. The probe was ³²Plabeled TAN1 cDNA (ANS) or rice DNA encoding ribosomal RNA (rRNA) for a load control.

Japan) (Fig. 3A). The hybridization pattern suggested the presence of a single-copy ANS gene in T. fournieri. Two apparent bands upon digestion by EcoRI and EcoRV are presumably due to the presence of these restriction sites in intron region or partial digestion of DNA.

For RNA blot analysis, total RNA was extracted using RNeasy Plant Mini Kit (Qiagen) from petals of flower bud and mature flower, and leaf, according to manufacture's instructions. Ten micrograms of RNA was separated under a denaturing condition on a 1.2 % agarose gel containing formaldehyde and then transferred onto a Hybond-N+ membrane. Hybridization of the membrane was performed by the same probe under the same condition for Southern blot analysis. To verify equivalent loading of RNA on the blot, the membrane was also probed with ³²P-labeled rice DNA for ribosomal RNA, pRR217 (Takaiwa *et al.*, 1985). As shown in **Fig. 3B**, ANS transcript (1.4 kb) was detected only in petals of flower bud and no evident signal of mRNA expression was observed in mature flower petal and leaf. This expression pattern is well consistent with the timing of coloring in petal of *T. fournieri*. The similar developmentally regulated expression of ANS gene was reported for the flower of *Forsythia* x *intermedia* (Rosati *et al.*, 1999).

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