

## Gene Note

## Isolation and expression analysis of phosphate transporter genes from *Eucalyptus camaldulensis*

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**Abstract** Five distinct genomic DNA fragments (*EcPT1*, *EcPT2*, *EcPT3*, *EcPT4* and *EcPT5*) encoding phosphate transporters were isolated from *Eucalyptus camaldulensis*. *EcPT2* and *EcPT3* were exclusively expressed in the root, but were not enhanced by phosphate deprivation. The transcript level of *EcPT2* was much higher than that of *EcPT3*. The *EcPT2* is present as a single copy gene in *Eucalyptus* genome, and phylogenetic analysis and deduced amino acid sequence revealed that *EcPT2* can be classified into the Pht1 family, a high-affinity phosphate transporter. These results suggest that *EcPT2* functions in phosphate transport in root tissues not only under low-phosphate conditions as with typical Pht1 family members, but also under high-phosphate conditions.

**Key words:** *Eucalyptus*, phosphate transporter, Pht1 family.

Phosphorus (P) is one of the most important nutrients for plant growth and, since it is deficient in most soils, limits plant growth in most natural ecosystems (Bielecki 1973). In agricultural systems, a large input of phosphate fertilizer is required since P is acquired by plants in the form of inorganic phosphate (Pi). Plants use a series of strategies to enhance Pi acquisition from soils, including secretion of several molecules (protons, organic acids, phosphatases and nucleases), changes in root morphology and up-regulation of Pi transporters (Poirier and Bucher 2002; Rausch and Bucher 2002). Of these strategies, it is likely that Pi transporters in roots play a major role in Pi uptake from the soil. Understanding the molecular mechanisms of Pi uptake is thought to be an important step in gene manipulation of high-yield plants in Pi-deficient soils.

Although Pi transport has been extensively investigated in various plants, including agriculturally important crops, little is known with regard to industrially important wood species. Recently, with the increasing demand for renewable energy, the importance of fiber and chemical materials from wood has been growing rapidly. Breeding of trees that can grow in nutritionally poor soil will expand the plantation area, increase wood production and reduce pressure of exploitation of native forests.

*Eucalyptus* is important as a raw material for

industrial pulp and paper making. In this study, as a first step in understanding the molecular mechanism of Pi uptake in *Eucalyptus*, we focused on the Pi transporter genes of this species.

Pi transporter genes were screened from the *E. camaldulensis* genomic library. A cDNA fragment encoding a putative Pi transporter derived from the *Eucalyptus* EST database constructed by Oji Paper Co. Ltd. (Japan) was used as a probe for plaque hybridization. Five partial genomic DNA sequences with high homology to the Pi transporter gene were isolated. These genes were designated *EcPT1* (identical to the EST sequence used as probe), *EcPT2*, *EcPT3*, *EcPT4* and *EcPT5*. Gene accession numbers are AB242816, AB242817, AB242818, AB242819 and AB242820, respectively. Each of these partial DNA sequences shares around 80% homology.

Expression of these five genes was analyzed by northern blot analysis. *EcPT2* and *EcPT3* mRNA were expressed in a root-specific manner and were not induced by Pi-deficiency (Figure 1). Moreover, *EcPT2* mRNA accumulation was much higher than that of *EcPT3*. Expression of *EcPT1*, *EcPT4* and *EcPT5* was not observed in the current experiment. Further, although *EcPT1* was observed in the EST database, the transcript of *EcPT1* was not detected. This might have been due to the low expression of *EcPT1* in the experimental

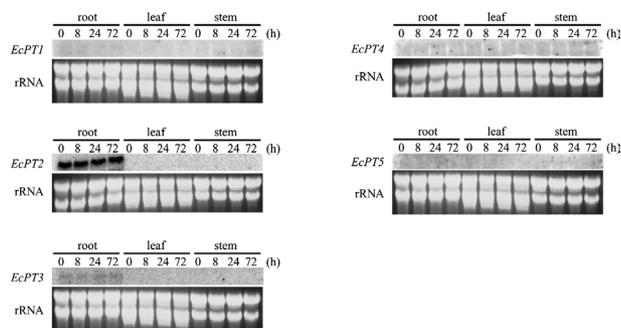


Figure 1. Expression analysis of the *EcPT* genes in *E. camaldulensis*. *Eucalyptus* plants with the same genetic background were used throughout this study. Plantlets were grown in 1/4 strength Gamborg's B5 medium solution supplemented with  $0.5 \text{ g l}^{-1}$  2-(*N*-Morpholino) ethanesulfonic (MES) (pH 5.6) for about 1 month in a greenhouse. To confirm the response to Pi deficiency, medium containing  $250 \mu\text{M}$   $\text{NaH}_2\text{PO}_4$  was replaced with Pi-deficient medium containing  $250 \mu\text{M}$   $\text{NaCl}$  0 h, 8 h, 24 h and 72 h before sampling. Total RNA was isolated from the leaves, stems and roots as described by Suzuki et al. (2003). Aliquots of  $15 \mu\text{g}$  of RNA were electrophoretically separated on formaldehyde agarose gel and blotted onto a nylon membrane. Hybridization was carried out according to a standard procedure. Details of the sequence information of the probes and primers used for amplification are available from the authors on request.

conditions used in this study.

We therefore focused on *EcPT2* since the expression level of this gene in the roots was extremely high. *EcPT2* has one intron within its coding region (Figure 2A). The deduced amino acid sequence of *EcPT2* consists of 535 amino acids with an apparent molecular mass of 58.4 kDa. Genomic Southern blot analysis showed the presence of a single *EcPT2* gene in *E. camaldulensis* (Figure 2B).

In vascular plants, genes encoding Pi transporters and Pi translocators (PT) play a crucial role in Pi transport systems. Recently, Pi transporters were functionally and structurally classified into three families, namely, Pht1, Pht2 and Pht3 (Rausch and Bucher 2002). Phylogenetic analysis of *EcPT2* with *Arabidopsis* Pi transporters/translocators demonstrated a close relationship between *EcPT2* and the Pht1 family (Figure 2C). Kinetic data for Pht1 members showed that they were high-affinity Pi transporters and involved in acquisition of Pi by the roots from low external concentrations in the soil (Daram et al. 1998; Mitsukawa et al. 1997). The TMAP program

(<http://bioinfo.limbo.ifm.liu.se/tmap/>) predicted the presence of 12 transmembrane domains in *EcPT2* and a central hydrophilic domain separating six N-terminal domains from six C-terminal domains (Figure 2D). This domain structure is common among other high-affinity Pi transporters of higher plants, yeast and fungi (Ming et al. 2005; Harrison et al. 2002; Paszkowski et al. 2002; Rausch et al. 2001; Muchhal et al. 1996; Harrison et al. 1995; Bun-Ya et al. 1991). The phosphorylation sites for protein kinase C and casein kinase II and the site for potential *N*-glycosylation are conserved in high-affinity Pi transporters (Ming et al. 2005; Kai et al. 2002; Muchhal et al. 1996) and are also present in *EcPT2* at positions 238–240 (T-A-R), 505–508 (S-L-E-E) and 419–422 (N-A-T-T), respectively.

To date, many genes encoding a high-affinity Pi transporter homologous to yeast *PHO84* have been identified in plants, helping elucidate Pi uptake mechanisms (Poirier and Bucher 2002; Rausch and Bucher 2002). Most are expressed predominantly in root tissues and are strongly induced by Pi deprivation. In *Arabidopsis*, all *Pht1* genes, except *Pht1;6*, are expressed either exclusively or predominantly in the roots, and are induced under conditions of Pi deprivation (Mudge et al. 2002). In rice, 10 of the 13 Pi transporters are expressed in the roots (Paszkowski et al. 2002). Root-specificity and Pi-deficiency inducible expression are typical features of *Pht1* family genes across various plant species (Ming et al. 2005; Schunmann et al. 2004; Chiou et al. 2001; Daram et al. 1998).

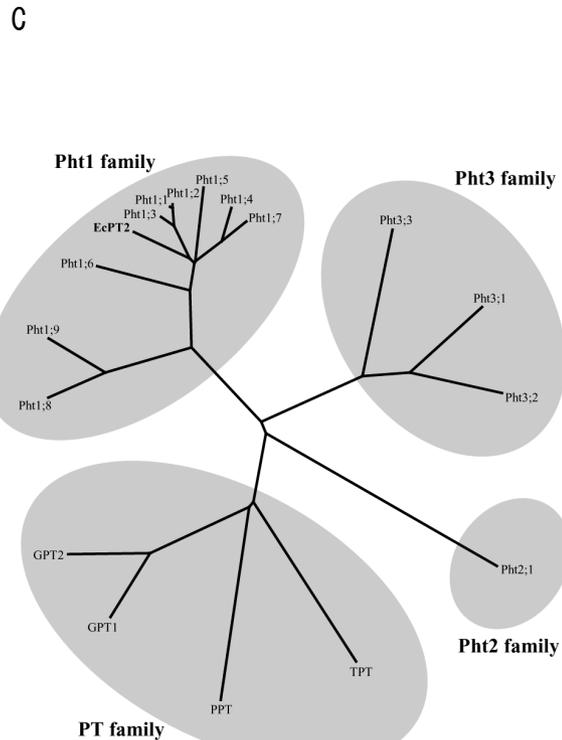
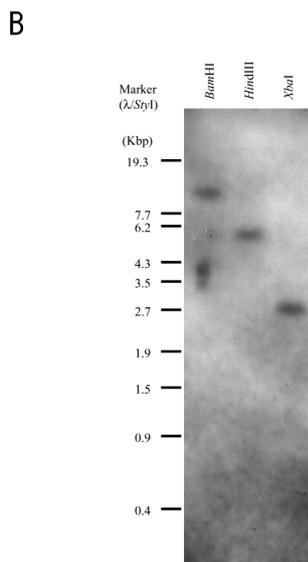
In this study, five putative Pi transporter genes were isolated from the genomic library of *E. camaldulensis*. Of these, only two, designated *EcPT2* and *EcPT3*, were expressed constitutively in a root-specific and Pi-independent manner in our experimental conditions. Nucleotide sequencing of the *EcPT2* implied that this gene encodes a high-affinity Pi transporter, suggesting that *EcPT2* functions as a Pi transport both in low and high Pi conditions. Isolation of a full set of *Pht1* genes and detailed expression studies of *EcPT2* are currently in progress with the aim of further understanding the Pi transport mechanism in *Eucalyptus*.

Figure 2. Properties of *EcPT2*. (A) Nucleic and deduced amino acid sequences of *EcPT2*. The deduced amino acid of the ORF is shown under the nucleic acid sequence. An intron (underlined) was presumed using a splicing site prediction program (NetGene2 Server; <http://www.cbs.dtu.dk/service/NetGene2/>). This splicing site was also confirmed by sequencing of *EcPT2* cDNA amplified from *Eucalyptus* root RNA by RT-PCR using gene-specific primers. (B) Southern blot analysis of *EcPT2*. Genomic DNA was extracted from *Eucalyptus* leaf as described by Wagner et al. (1987) with minor modifications. Aliquots of  $15 \mu\text{g}$  of genomic DNA were digested with the indicated restriction enzymes, electrophoretically separated on agarose gel and blotted onto a nylon membrane. DIG-labeled (Roche, Germany) *EcPT2* DNA (+1086 to +1609 relative to the translational start site with no restriction enzyme site used for the genomic DNA digestion) was used as a probe. Hybridization was carried out according to a standard procedure. (C) The phylogenetic tree of *EcPT2* and *Arabidopsis* Pi transporter/translocator proteins. Amino acid sequences were aligned using ClustalW and the tree was constructed with TreeView software. (D) Amino acid sequences of *EcPT2* and its related Pi transporter proteins. The deduced amino acid sequence of proteins *EcPT2*, *Pht1;1*, *Pht1;4* and *Pht1;6* were aligned. The predicted transmembrane (TM) domains of *EcPT2* are indicated by a dotted line. The phosphorylation sites of protein kinase C (\*) and casein kinase II (#) and the site of potential *N*-glycosylation (+) are indicated.

**A**

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1 atgctaag aaatottg ggtotaat acactgatg tggcoagac acaatgtao
M A K E N L G V L N A L D V A K T O W Y
61 catttaooc coactatoc tgcogcoag eegttotica ocgacogta tcatotctt
H F T A I I I A G M G F F T D A Y D L F
121 tgcatttoco tegtocagaa gctocogt cgcatact acatgagga taagataag
C V S L V T K L L G R I Y Y Y E G K D K
181 cctgctgc tooctocga cgtgctgoc gctgcoagc gctgoccti ctgogtact
P G S L P P N V A A A V N G V A F C G T
241 ctacogcogc agctotctt cggctgott gggacaat tagtoggaa gogtgotat
L A G O L F F G W L G D K L G R K R V Y
301 ggttaooc toattat gacatogc toctogct cggctgto cttgggat
G L T L L I M I I C S V G S G L S F G D
361 tcocogaata gtttatagc aaoctatgc ttctocgat tctgctgoc gttgggato
S P N S V I A T L C F F R F W L G F G I
421 ggagggatt accoactoc cocacoato atgogagat agcogaaca gaagactogt
G G D Y P L S A T I M S E Y A N K K T R
481 ggcoattoa toctogcag tttgocagc caaggtttg ggtttggc ggtgggato
G A F I A A V F A M O G F G I L G G G I
541 gtcottga tttotococ agactogt cataagttta agccocogc ttagaagtc
V A L I V S S A F D H K F K A P P Y E V
601 aatocagtg gctogcagc toctocagc gattagctg gogtatcat tgaatgoc
N P V G S T V P Q A D Y V W R I I V M L
661 ggtcoattac cocogococ tactattat tatagaaga aatgocgta aactgocgt
G A L P A A L T Y Y Y R M K M P E T A R
721 tacacogcc toctogcaag aaacggaag caggagctg cagacatgc taagtaga
Y T A L V A R N G K Q A A A D M S K
781 acaagata gtttotoct tgcocataa acaatcact agttattgg agaattata
841 gatgatac caagactata atctgactt attagatgg cctocaga attotittg
901 aactgctot toctittga gtoactgac atagttagg tttttotato aaattattg
961 gtttgcatt cttgcoata cttttotgt tagtgcaa tttcatgta ttaatagaa
1021 accgtaaca gttttgcaa gttgatgg aatcagaaca agagaagtg gagaattta
V L Q V D I E S E Q E K V E K F I
1081 cgaagatoc cocoacagc tagtgcott toctogaaga atcogocogc ogcoagcogc
O D P R N S Y G L F S K E F A R R H G L
1141 toacactogt gggacagoc acoactggt toctogoga catogottto tatoccaaa
H L V G T A T T W F L L D I A F Y S Q N
1201 acototcoa aaagatatt ttoacogca toctggctgt ccagocogcc aagaagata
L F O K D I F T A I G W L P A A K K M N
1261 agcoattoa ogagatac aatattgca ggcocaaac cctgatogct ctttgagta
A I H E V Y K I A R A O T L I A L C S T
1321 ctgctogtg gttattgto actgocoga ogatogatta catogaga tocttoatc
V P G Y W F T V A T I D Y I G R F F I O
1381 aatgatgg tttogcagc atgcaatct toattttgc catogocato coatacaac
M M G F A M M S I F M F A I A I P Y N H
1441 actgaaca toacatata gattogctg ttagtactc actcaocto toctogca
W K H H H I G F V V M Y S L T F F F A N
1501 acttggoc aaocogaaca aactcattg ttoocogta gatottocog gogcgtta
F G P N A T T F I V P A E I F P A R L R
1561 gatocactg cactgocata tocgocctc oaggaagc ttagacact gttggcgtt
S T C H G I S A A A G K A G A I V G A F
1621 tttgcttct gtagcogc cagataaaa caagocoga tgcagttat aocogcga
G F L Y A A Q D K T S P D A G V K P G I
1681 tocgogtaa gaatgocgt ctigtcttg gggagtaa toctogoga atgtttoa
G V K N A L L V L G G I N L A G M L F T
1741 ctttctgt gcccagaca aagcagct cactgagga aattgagga gagaacatg
L L V P E P K G R S L E E I G G E N M D
1801 atgataaga gatggtaga gtaggagca tgcocotoc ttoctggca coatttgg
D N E M G E G R E M Q P P S L G P F G G
1861 gttag
    
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**D**

EcPT2	1	WAKENLGVLNALDVAKTONYHFTATIIAGMGFFTDAYDLFCVSLVTKLLGRYYYYEKG-	TM1
Pht1.1	1	WAEQQLGLVLLKALDVAKTONYHFTAVIIVAGMGFFTDAYDLFCVSLVTKLLGRYYYYEKS	
Pht1.4	1	WAEQQLGLVLLKALDVAKTONYHFTATIIAGMGFFTDAYDLFCVSLVTKLLGRYYYYEKA	
Pht1.6	1	WAEQQLGSLVLLKALDVAKTONYHFTAVIIVAGMGFFTDSDYDLFVLSLTKLLGRYYYYEKG	
EcPT2	59	DKPGSLPPNYAAAVNGVAFVCGTLGQLFFGWLGDKLRKRYVYGLTLLINTICSVGSLSL	TM2
Pht1.1	60	AKPGSLPPNYAAAVNGVAFVCGTLGQLFFGWLGDKLRKRYVYGLTLLVMMILCSVASLSL	TM3
Pht1.4	60	QKPGILPPNYAAAVNGVAFVCGTLGQLFFGWLGDKLRKRYVYGLTLLVMMVLCISLASLSL	
Pht1.6	61	SSPGSLPPDYSAAVSVGVAFAVGTITGQIFFGWLGDKLRKRYVYGLTLLINTICSVGSLSL	
EcPT2	119	GDSRNSVIAALGFFRFLWFGFVGGDYPLSATIMSEYANKKTRGAFIAAVFANQGGFLLGG	TM4
Pht1.1	120	GHEAKGVMITLGFRRFLWFGFVGGDYPLSATIMSEYANKKTRGAFIAAVFANQGGVILGG	
Pht1.4	120	GHEPKAVMATLGFRRFLWFGFVGGDYPLSATIMSEYANKKTRGAFIAAVFANQGGIJJGG	
Pht1.6	121	GRDFKTYMVTLGFRRFLWFGFVGGDYPLSATIMSEYANKKTRGAFIAAVFANQGGVILAA	
EcPT2	179	GIVALVSSAFDHKKAPPYEVNPGVSYPOADYYVRIIVMLGALPAALTYWRNKMPET	TM5
Pht1.1	180	GFVALVSSAFEDKKEPAPTYAVNRALSTPOADYYVRIIVMFGALPAALTYWRNKMPET	TM6
Pht1.4	180	GTFALVSSAFEAKEFSPAYADALGSLPOADYYVRIIVMAGALPAALTYWRNKMPET	
Pht1.6	181	GAVSLVSAVVFESKFPNAYILDGAASTYPOADYYVRIIVMFGALPAALTYWRNKMPET	
EcPT2	239	ARYTALVARNGKQAAADMSKVLQVDTESEGEKVEKFTQDPRNSYGLFSKFAARRHGLHLV	
Pht1.1	240	ARYTALYAKNIKQATADMSKVLQVDTELEERVEDDVKDPKONYGLFSKFAARRHGLHLI	
Pht1.4	240	ARYTALYAKDAKQAAADMSKVLQVDTEPEQOKLEIEISKESKAFGLFSKFAARRHGLHLI	
Pht1.6	241	ARYTALVSKNAQAAADMSKVLQVDTEASAANKDQ-ARVSDDEGLFSKFAARRHGLHLI	
EcPT2	299	GATITWFLLDIAFYSONLFQKDIPTAIGWLPAAKKNWATHEVYKIAQAOTLLALCSTVPG	TM7
Pht1.1	299	GTTSTWFLLDIAFYSONLFQKDIPTAIGWLPKAAIWNATHEVERIAQAOTLLALCSTVPG	
Pht1.4	300	GTTSTWFLLDIAFYSONLFQKDIPTAIGWLPAAQSNWATHEVYKIAQAOTLLALCSTVPG	
Pht1.6	300	GATSTWFLLDIAFYSONLFQKDIPTIIGWLPAAKKNWATHEVYKIAQAOTLLALCSTVPG	
EcPT2	359	YWFIVAFIDYIGRFKIQMMGFAMMSDFMFAIATPYNHWK---HHHIGLVVMYSLTFFFAVF	TM8
Pht1.1	359	YWFIVAFIDYIGRFKIQENGFVMMIVMFAIATPYNHWK---HHHIGLVVMYSLTFFFAVF	TM9
Pht1.4	360	YWFIVAFIDYIGRFKIQMMGFAMMSDFMFAIATPYNHWK---HHHIGLVVMYSLTFFFAVF	
Pht1.6	360	YWFIVAFIDYIGRFKIQMMGFAMMSDFMFAIATPYNHWK---HHHIGLVVMYSLTFFFAVF	
EcPT2	417	GNATTFVPAEIPPARLRSTCHGISAAAGKAGAVGAFGLYAAQ---DKTSPDAGYKPG	TM10
Pht1.1	419	GNATTFVPAEIPPARLRSTCHGISAAAGKAGAVGAFGLYAAQSDOKAVVDAGYPPG	TM11
Pht1.4	420	GNATTFVPAEIPPARLRSTCHGISAAAGKAGAVGAFGLYAAQNDKAVVDAGYPPG	
Pht1.6	420	GNATTFVPAEIPPARLRSTCHGISAAAGKAGAVGAFGLYAAQNDKAVVDAGYPPG	
EcPT2	475	IGVKNALVVLGGTLLAGMLFTLLVPEPKGRSLEEIGGENMDDNEGEGREIMOPPSLGGPF	TM12
Pht1.1	479	IGVKNSLVLLGVLMFVIGMLFTLLVPEPKGRSLEEISG---EAEVSHDEK-----	
Pht1.4	480	IGVKNSLVLLGVLMFVIGMLFTLLVPEPKGRSLEEISG---EDNEVSHNDSRTVGLV---	
Pht1.6	467	IGVSNLTYLMAGINLLGLLFTLTPETPKGRSLEEISG---EIEPEKIKKIKV---	
EcPT2	535	G	###
Pht1.1	-	-	
Pht1.4	-	-	
Pht1.6	-	-	

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